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Molecular characterization of fowl and pigeon pox viruses: A recent outbreak in Libya

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Abstract

Background: Poxvirus infections in poultry, particularly fowl poxviruses (FPV), pose significant challenges to the global poultry industry. A notable outbreak of cutaneous fowl (FP) and pigeon pox in Libya has primarily affected backyard chickens, pigeons, and some commercial layers, marking the region's first official record of these viruses.

Aim: This study aimed to isolate, identify, and characterize FP and pigeon pox viruses (PPVs) associated with the outbreak.

Methods: Histopathological examinations were conducted alongside clinical observations of lesions in affected chickens and pigeons. Virus isolation was performed using embryonated chicken eggs, and molecular diagnosis was achieved via polymerase chain reaction targeting the P4b core protein gene. Sequencing and phylogenetic analyses were also performed.

Results: Characteristic lesions, such as wart-like growths and scabs, were observed in backyard chickens and pigeons. Histopathological analyses confirmed the presence of eosinophilic intracytoplasmic inclusion bodies. Molecular analysis revealed high genetic similarity among the four FPV isolates, which were 100% identical to selected isolates from Iraq, Iran, and Brazil. The PPV isolates also showed significant genetic homogeneity, with 100% identity to strains from Egypt and India and high similarities to other isolates.

Conclusion: Our findings underscore the need for further investigation into the epidemiology and transmission dynamics of fowlpox and PPVs. Future research should focus on the genetic diversity of PPVs and their implications for pathogenicity and host specificity. Ongoing global surveillance and genetic analysis of avian viruses are crucial for understanding their impact on poultry populations and developing effective disease management strategies.

Keywords: Fowl pox, Pigeon pox, Backyard chicken, Pigeon, Libya.

Introduction

The genus Avipox encompasses several DNA viruses, including Fowlpox virus (FPV), Pigeon Pox virus (PPV), Turkey Pox virus, Canary Pox virus, and Penguin Pox virus. Fowl pox (FP), a virus of this genus within the Poxviridae family, is a globally distributed and slowly spreading viral disease affecting chickens. All bird species are vulnerable to avian pox, as evidenced by reported cases of natural infections in various wild and domestic birds (Fauquet *et al.*, 2005). In contrast to DNA viruses, poxviruses replicate and develop in the cytoplasm of infected cells. The transmission of pox infections is significantly facilitated by wild birds and insects. The FPV has a DNA structure of approximately 288 to 300 kilobase pairs in length (Asif *et al.*, 2021). Characteristically, poxviruses possess a distinctive brick-like shape and spread slowly, leading to the development of proliferative lesions and scabs (dry form) on the skin, along with diphtheritic lesions (wet form) in the upper

digestive and respiratory systems (Williams *et al.*, 2021).

FP has considerable economic implications because it can decrease egg production and hinder growth in younger birds. The risk of mortality increases when both dry and wet forms are present (Singh *et al.*, 2000). Typical clinical symptoms include weight loss, feather loss, and scaly skin on the head, neck, and back. Secondary bacterial infections are common in both forms and can result in pneumonia or further infections at blister locations (Tripathy and Reed, 2013). Prompt recognition of these clinical signs, timely intervention, and appropriate management practices are crucial for mitigating the impact of poxvirus outbreaks on poultry health and welfare. Veterinarian consultation and diagnostic testing are essential for confirming poxvirus infections and implementing targeted control measures. Preventing poxvirus outbreaks in poultry requires a multifaceted approach that incorporates robust biosecurity measures, vaccination programs, hygiene protocols, and effective farm management practices.

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By integrating these practices into daily operations and continuously monitoring strategies to meet the evolving needs of the flock, poultry farmers can significantly reduce the risk of poxvirus outbreaks and enhance the health and well-being of their poultry populations. Collaborations with veterinary authorities, industry experts, and peers can further bolster disease prevention efforts and strengthen the overall biosecurity of poultry production systems (Giotis and Skinner, 2019).

In Libya, there is significant interest in backyard chickens and pigeons, yet no official reports exist on poxvirus infections, and data on clinical cases from veterinarians are scarce despite observed cases in the field. The Libyan Animal Health Authority recommends vaccinating against poxvirus at 4–7 weeks of age for protection against poxvirus. However, the lack of recorded data on FP and pigeon pox, along with limited characterization of the viruses, leaves significant gaps in our understanding of these diseases. This study is the first official documentation of fowl and PPVs in Libya, utilizing isolation, identification, and genetic and phylogenetic analyses.

Materials and Methods

Sampling

Between January 2022 and December 2022, cases submitted to the Faculty of Veterinary Medicine Hospital revealed pox lesions in commercial chickens, backyard chickens, and pigeons. These cases were observed and humanely euthanized, and skin samples were collected from the lesions. The samples were divided into two portions: one was preserved in 10% buffered formalin, and the other was stored in a deep freezer for egg inoculation.

Additionally, older frozen samples were obtained from private veterinary clinics that had stored them in a deep freezer since 2014. These included samples from feral pigeons collected in the Tripoli area, as well as samples from backyard chickens gathered in 2016 and 2017 from the Souq Aljuma and Janzour areas of Tripoli, respectively. Another sample of the pigeon species, *Columba rupestris*, was collected in 2019 from

the Tajoura area in Tripoli. All frozen samples were thawed, and the material from the pox lesions was used for egg inoculation. Table 1 summarizes the samples received during this period.

Histopathology

Sections of skin with pox lesions were cut into pieces, including all layers of the skin, from samples from the current study. The skin samples were preserved in neutral formalin until processing. After being washed overnight in running water and dehydrated in increasing concentrations of alcohol, the tissues were embedded in paraffin. Sections 5 µm thick were then cut and stained with hematoxylin and eosin.

Methods of egg inoculation

The methods used for chorioallantoic membrane (CAM) inoculation were described in (Baddeley and Tripathy, 1998) with minor modifications. Fertile eggs were obtained from flocks under restricted control and observation, in which previously does not receive any type of vaccination during the entire period of their lives.

polymerase chain reaction (PCR)

The QIAamp DNA Mini Kit. Oligonucleotide primers used in conventional polymerase chain reaction (Midland Certified Reagent Company_ oilgos, USA). Oligonucleotide primer sequences were used according to (Prukner-Radovčić et al., 2006) with amplified product 578 bp of gene 4b core primer Sequence (5'-3'). The extraction of viral DNA was conducted according to QIAamp DNA mini kit instructions. Briefly, each cycle of the reaction consisted of primary denaturation at 94°C for 2 minutes, secondary denaturation at 94°C for 1 minute, annealing at 60°C for 1 minute, and extension at 72°C for 1 minute. The cycle was repeated 35 times with a final extension at 72°C for 2 minutes. The gel was then photographed by a gel documentation system. A purified PCR product was sequenced in the forward and/or reverse directions on an Applied Biosystems 3130 automated DNA Sequencer (ABI, 3130, USA) using a cycle sequencing kit (Perkin-Elmer/Applied Biosystems, Foster City, CA).

Table 1. Samples collected during the studies indicating the type and breed of birds and location.

No.	ID	Host	Breed	Type of sample	Time of sample collection	Location
1	LM	Chicken	Novo gen white	Skin tissue	12–2022	Wady Elrabi, Tripoli's
2	LT	Pigeon	<i>Columba rupestris</i> .	Skin tissue	8–2019	Tajora, Tripoli
3	BM	Chicken	Back yard	Skin tissue	11–2016	Souq Aljuma,
4	BT	Chicken	Back yard	Skin tissue	1–2017	Janzor, Tripoli
5	LP	Pigeon	Feral pigeons	Skin tissue	8–2014	Tripoli Center
6	Li	Pigeon	White Doves of Wartburg Castle	Skin tissue	12–2022	Tripoli Center
7	IW	Chicken	Hy-line W- 80: white	Skin tissue	12–2022	Gaser Benghashir,
8	LW	Pigeon	Duchesse Utility Pigeon	Skin tissue	12–2022	Elfornaj, Tripoli



Fig. 1. Commercial Novo gen white-laying chickens aged 18 weeks with pox lesions on their combs and wattles.



Fig. 2. Commercial laying chicken (Hy-line W- 80 white) aged 16 weeks with a wart-like lesion on the tops of their peaks.

The Basic Local Alignment Search Tool (A BLAST® analysis) was initially used to establish sequence identity for GenBank accessions. The sequence reaction was performed according to the instructions of the manufacturer.

Phylogenetic analysis

A comparative sequence analysis was conducted using the CLUSTAL W multiple sequence alignment program, version 12.1 of the MegAlign module from LaserGene DNASTar software (Madison, Wisconsin, USA), developed by (Tamura *et al.*, 2013). Phylogenetic analyses were performed using maximum likelihood neighbor joining (MEGA11 software).

Ethical approval

This research was approved by the Scientific Research and Ethics Committee of the University of Tripoli (No.: SREC/010/33) dated on 31/07/2023.

Results

Clinical signs and lesions

Samples included in the current study exhibited Pox lesions consisting of a cutaneous form (dry pox) characterized by the growth of proliferative lesions, which varied from small nodules to spherical wart-like masses on the skin of the comb, wattle, and other featherless areas. The samples belonged to Novogen chickens (Fig. 1), commercial chickens (Fig. 2), and backyard chickens (Fig. 3). In addition, four samples from a private clinic were frozen from feral pigeons since 2014. Additionally, a sample of a *C. rupestris* pigeon was frozen in 2019. Two samples from backyard chickens were also frozen, one in 2016 and the other in 2017.

Histopathology of skin samples

Only fresh chicken and pigeon samples were sent for histopathological analysis. Histologic sections of the skin from areas over the eye, combs, wattles, and around



Fig. 3. Backyard chicken aged 40 days suffered from severe pox lesions on the comb, wattles, and around the eyes.

the mouth exhibited significant epithelial hyperplasia, resulting in deep folds. Some differences were noted in the extent of severity of histopathological lesions among the samples included in the current study.

The skin of the backyard chicken showed multifocal hyperplasia of the epidermal epithelium with mild ballooning degeneration (swollen) of keratocyte and numerous eosinophilic viral inclusion bodies (Bollinger bodies) within the cytoplasm of the affected cells. A section is characterized by the presence of a moderate amount of superficial amorphous eosinophilic necrotic material, admixed with cellular

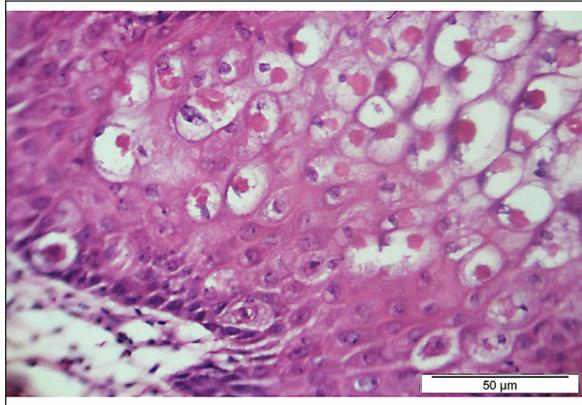


Fig. 4. Section of sample showing extensive lesions (ballooning degeneration and inclusion bodies) caused by PPV in the skin of *C. rupestris* pigeons (HE, Bar = 50 µm).

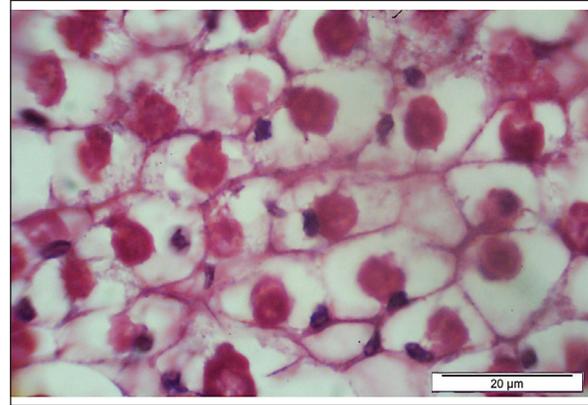


Fig. 5. Section of sample showing Bollinger bodies displacing the nuclei of keratocyte to the periphery (HE, 20 µm).

debris. Whereas in sample skin of backyard chicken and sample skin of pigeon, the epidermal and follicular epithelium is diffusely hyperplastic with marked thickness of the stratum spinosum that shows extensive swollen keratinocytes and the presence of eosinophilic viral inclusion bodies within the cytoplasm of the affected cells displacing the nuclei to the periphery (Figs. 4 and 5).

Virus isolation

Samples of live birds and frozen samples were used for egg inoculation to propagate the virus. Three passages were conducted on the eggs. All isolates grew well on the CAM. Pock lesions were clear on CAMs inoculated with samples from backyard chickens and commercial chickens (Fig. 6). However, isolates collected from pigeons did not exhibit clear pock lesions on the CAM, whereas all samples showed thickening of the CAM accompanied by varying degrees of hemorrhage. The results of egg inoculation for each isolate are presented in Table 2.

Histopathology of the CAM samples

The samples showed differences in the severity of the reaction, especially in the presence of intracytoplasmic inclusion bodies, as shown in Table 3. The histopathological lesions observed on CAM inoculated from samples from backyard chickens were thickening of the CAM tissue with infiltration of lymphocytes, presence of syncytial cells and hemorrhages (Fig. 7), presence of intracytoplasmic eosinophilic inclusion bodies, and ballooning of some other cells (Fig. 8).

PCR

The PCR results are shown in Figure 9. All eight samples included in the current study were positive for the pox virus using conventional PCR. The 578 bp of 4 b gene fragments (fpv167 locus) were amplified according to Lüscho *et al.* (2004).

Sequencing and phylogenetic analyses

Sequencing and phylogenetic analysis of the isolates from chickens and pigeons were performed. The

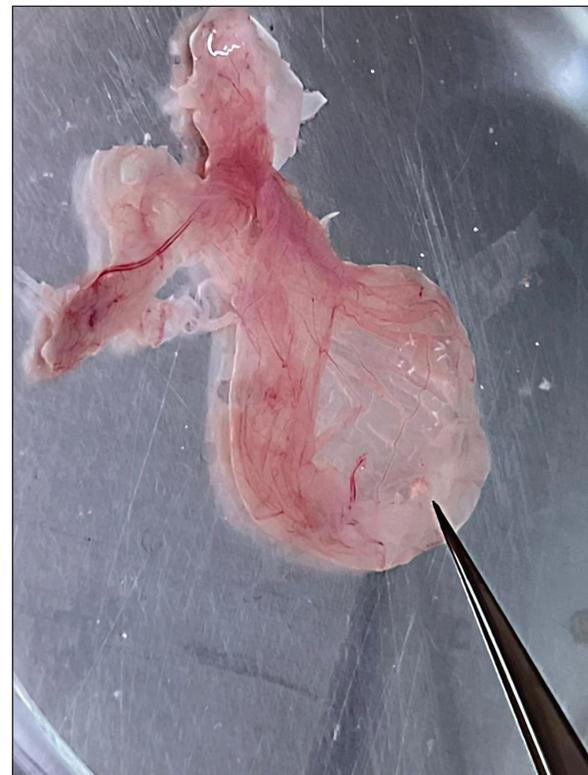


Fig. 6. Section of CAMs harvested from embryonated eggs inoculated with samples from backyard chickens. CAMs showing a membrane, hemorrhage, and pock lesions.

sequences were published in GenBank using the access numbers and names listed in Table 4. Partial sequences of the 4b core protein gene were amplified from four FP and four pigeon virus isolates. The obtained sequences (FASTA files) were subjected to NCBI BLASTN analysis to determine their identity with other isolates in the NCBI data archive. Phylogenetic analyses were

Table 2. Samples collected from the third passage on CAM after 5 days.

Sample NO.	Isolate NO.	Hemorrhages and thickening of CAM	POCK lesion in the CAM
1	Layer chicken (2022)	+	+
2	Pigeon (2019)	+	-
3	Backyard chicken (2016) (frozen sample)	+++	+
4	Backyard chicken (2017) (frozen sample)	+++	+
5	Pigeon (2014) (frozen sample)	+	-
6	Pigeon (2022)	+	-
7	Layer chicken (2022)	++	+
8	Pigeon (2022)	+	-

*(-) Negative, (+) few, (++) moderate, (+++) severe.

Table 3. the presence and intensity of cytoplasmic inclusion bodies in CAM inoculated with different isolates.

Sample No.	Isolate name	Presence of intracytoplasmic inclusion bodies
1	Layer chicken (2022)	+
2	Pigeon (2019) (frozen sample)	+
3	Backyard chicken (2016) (frozen sample)	+++
4	Backyard chicken, (2017) (frozen sample)	+++
5	Pigeon (2014) (frozen sample)	+
6	Pigeon (2022)	+
7	Chicken lying (2022)	+
8	Pigeon (2022)	+

*(-) Negative, (+) few, (++) moderate, (+++) severe

performed using maximum likelihood, neighbor-joining, and maximum parsimony in MEGA11 software, as shown in Figure 10. The four FPV isolates of the current study were very similar to each other, and the four pigeon virus isolates were also very similar to each other.

The FPV isolated in the current study were 100% identical to some selected isolates from Iraq, Iran, India, Brazil, South Africa, Gabon, and Egypt (Table 5). The similarities of the PPVs isolated in this study

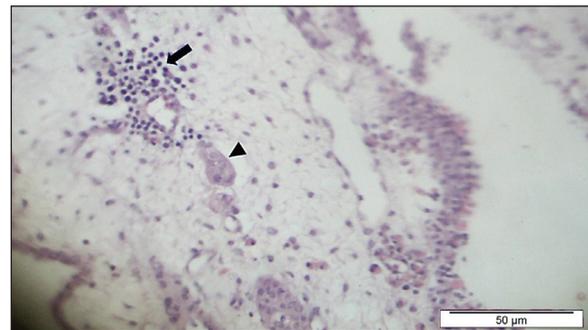


Fig. 7. Section of the CAM showing thickening of the tissue with infiltration of lymphocytes (arrow), presence of syncytial cells (arrow head) and hemorrhages (HE, Bar = 50 μm).

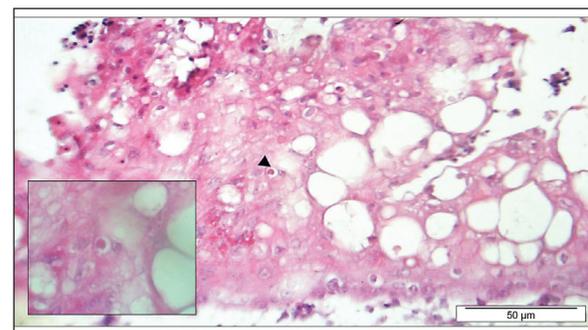


Fig. 8. Section of the CAM showing some infected cells (Arrowhead) with intracytoplasmic eosinophilic inclusion bodies and ballooning of other cells (HE, Bar = 50 μm, inset 100×).

are shown in Table 6, in which 100% similarities were found to isolates from Egypt and India. Identities of

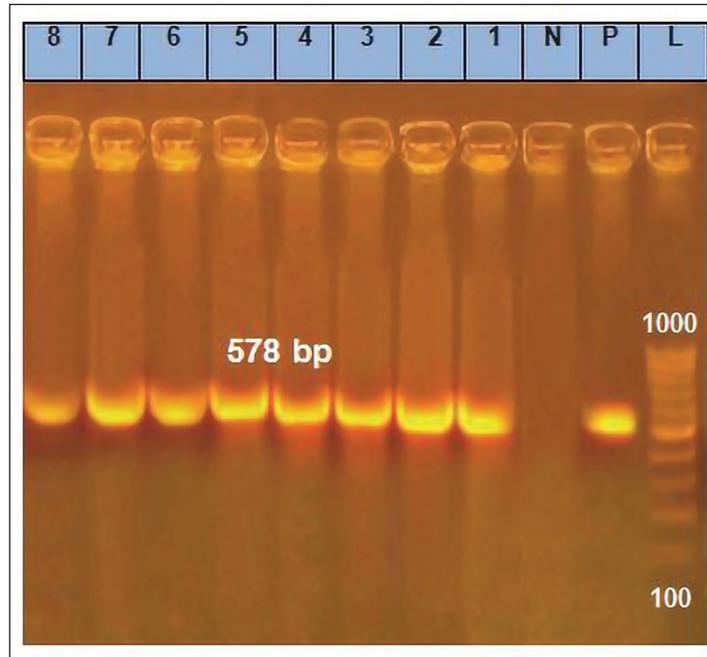


Fig. 9. Result of PCR showing the positive eight isolates from CAM. Bands of successful amplification of viral DNA (578 bp) compared with the positive and negative controls. A 1,000 bp DNA marker (Ladder) was used to determine the molecular weight of the bands.

Table 4. Sequencing data of poxviruses isolated from chickens and pigeons published in GenBank.

Sample no.	Access number	Full name	Bird species
1	OR099889	FPV strain LM core protein P4b gene	chicken
2	OR099890	FPV strain BM core protein P4b gene	Backyard chicken
3	OR099891	FPV strain BT core protein P4b gene	Backyard chicken
4	OR099892	FPV strain IW core protein P4b gene	Hy-line w-80, white chicken
5	OR099893	PPV strain LT core protein P4b gene	<i>Columba rupestris</i> (pigeon)
6	OR099894	PPV strain LP core protein P4b gene	Feral pigeon
7	OR099895	PPV strain Li core protein P4b gene	White dove
8	OR099896	PPV strain LW core protein P4b gene	Duchesse utility pigeon

99.80%, 99.78%, and 99.76% were isolated from Taiwan, India, and Iraq, respectively. Moreover, our isolates were 99.61% identical to other isolates from Egypt, Iraq, Iran, and Gabon, whereas our isolates were 99.60% identical to one isolate from Canada.

Discussion

Poxvirus infections in poultry have been documented for many years (Minhas *et al.*, 2024). Despite being a recognized disease in poultry, FPV continue to challenge the global poultry industry. Clinical manifestations of poxvirus infections in chickens and pigeons can be diagnosed by observing characteristic gross and microscopic lesions (Tripathy and Reed,

2013). The results of this study indicate that poxvirus infections in backyard chickens result in the formation of scabs on the comb, wattles, eyelids, and feet, consistent with the lesions described by Minhas *et al.* (2024). However, the lesions observed in commercial laying hens in the current study were atypical; they presented as wart-like lesions solely on the beak near the nasal orifice, with no lesions on the feathered areas. Similar atypical lesions have been previously reported in vaccinated commercial laying hen flocks in Brazil (Chacón *et al.*, 2020).

In the current study, backyard chickens and pigeons had more severe lesions, including small, bleeding, wart-like growths on areas such as the face, head, neck,

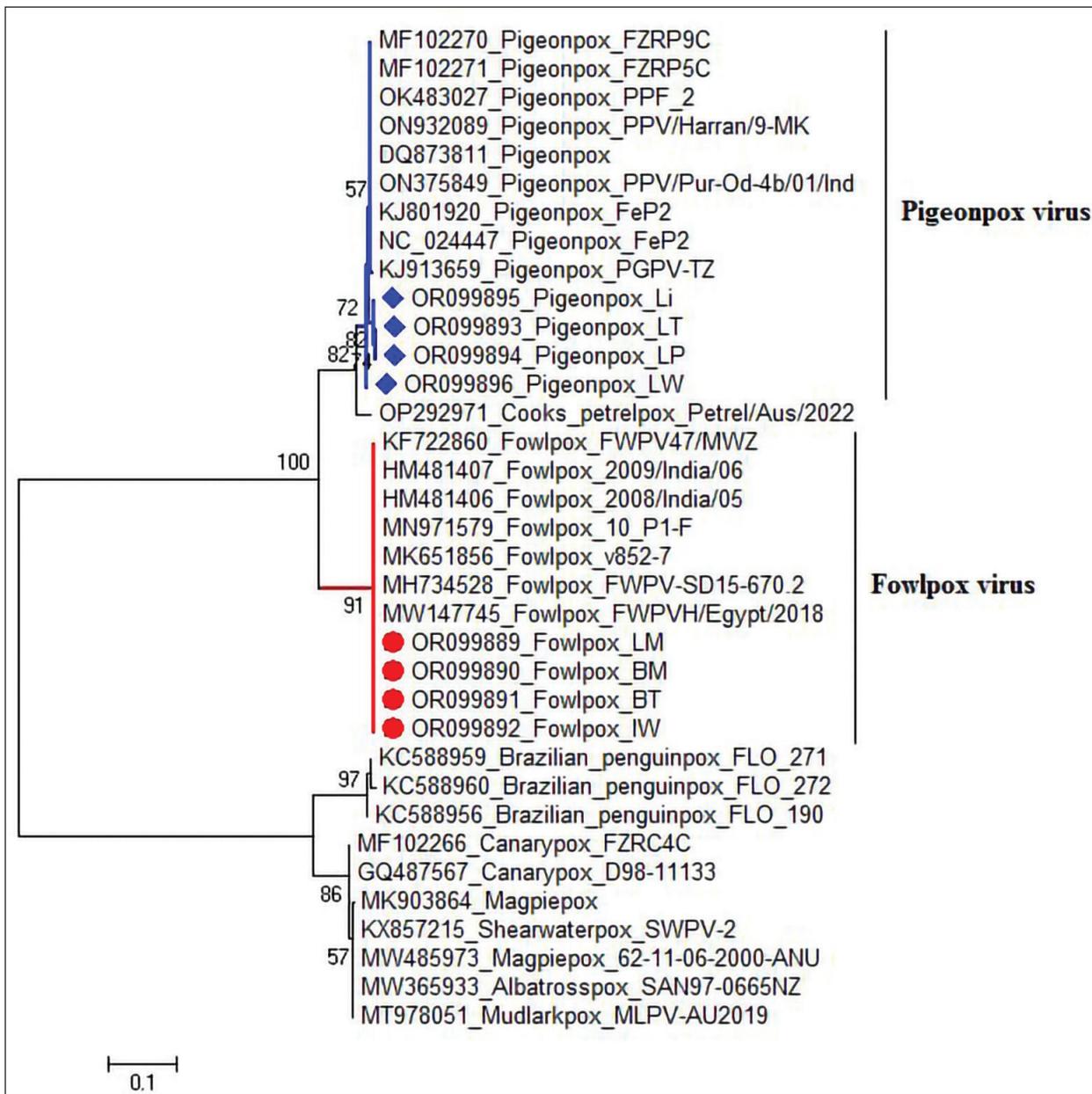


Fig. 10. Phylogenetic analysis of the partial 4b core protein gene of avian pox virus isolates. Isolate names and GenBank accession numbers. Libyan FP isolates from this study are highlighted in red circles, whereas Libyan pigeon isolates are highlighted in blue squares. The phylogenetic tree was constructed in MEGA 11 using the neighbor-joining method with 1,000 bootstrap replicates.

beak, and toes. These findings are similar to those described by Hibl *et al.* (2019). The differences in the severity of lesions among backyard chickens, pigeons, and commercial laying hens may be due to the lack of vaccination in the backyard chicken and pigeon populations.

Histopathological examination of fresh samples from backyard chickens, commercial layers, and pigeons demonstrates the presence of multiple eosinophilic intracytoplasmic inclusion bodies (Bollinger bodies),

which are a diagnostic feature of avian poxvirus infection (Tripathy and Reed, 2013). The epidermis showed marked hyperplasia (acanthosis) due to the swelling and increased number of cells in the stratum spinosum. These cells exhibited ballooning degeneration and contained various sizes of pale eosinophilic inclusion bodies, which are characteristic of avian pox (Tripathy *et al.*, 2000). Pustular lesions showed superficial crusts of hemorrhages and necrotic cells, with lymphocyte and histiocyte infiltration in

Table 5. Comparison of Libyan FP isolates with those from other populations (BLASTN).

Identity (%)	Origin of the isolate (country/city)	Year of submission	GenBank/Accession number	Isolate definition
100	Iraq: Salah Al Deen	2020	MN971579	FPV isolates the 10_P1-F 4b core protein gene
100	Brazil/Sao Paulo	2019	MK651857	FPV strain v852-8 4b core protein gene
100	India and Tamil Nadu	2018	MH447532	FPV isolates the PDDSL013 core protein (P4b) gene
100	Brazil/Caxias do Sul	2018	MG601782	FPV isolates the DF2016 4b core protein gene
100	India, Jammu and Kashmir	2017	MF496042	FPV major core protein P4b gene
100	Egypt and Giza:	2023	OR027043	FPV isolates the FP/EG/ISMAILIA-6 P4B gene
100	Iran/Lorestan	2015	KU212807	FPV isolates the LNHF-1 4b core protein gene and partial CDs
100	Gabon, Haut-Ogooue	2022	OQ064752	FPV isolate 2 4b core protein gene
100	South Africa, Gauteng	2015	KP987207	FPV isolate NGA_VRD09-172 P4b core protein gene
100	India – Kolkata	2021	MZ614741	FPV isolates the FPV/TRP/2021 4b core protein gene and partial CDs
100	Brazil / Rio de Janeiro	2021	OL703789	FPV isolates Cx A3334 core protein 4b gene
100	Austria–Vietnam	2021	OK345040	FPV isolates PA19/1236 (partial genome)

the dermis. Occasionally, fibroblast proliferation and fibrosis were observed in the affected areas.

In this study, the initial virus isolation was successful using 9-11-day-old embryonated chicken eggs (ECE) for inoculation of the isolate from chickens or pigeons, according to Tripathy and Reed (2013). Moreover, frozen old samples from 2014 to 2019 from pigeons and backyard chickens were readily grown and propagated in ECE, showing the typical histopathological changes of chicken embryos. These findings were similar to those of the studies by Offerman *et al.* (2013), which explained the hypertrophy and the appearance of many intracytoplasmic inclusion bodies in the ectoderm.

Three serial passages were continued for the propagation of FPV and PPV, as previously described (Sultana *et al.*, 2019). The CAM thickened upon inoculation with FPV and PPV samples, consistent with the findings of Kabir *et al.* (2015), Masola *et al.* (2015), and Roy *et al.* (2013). However, pock lesions on the CAM were absent in the pigeon samples. These findings are similar to the results reported (Audarya *et al.*, 2018). The results here agreed with earlier studies, except for the low number of pock lesions in most of the samples tested.

The P4b core protein gene is a unique genetic marker of poxviruses (Sarker *et al.*, 2021). The highly conserved characteristics of this gene enable the molecular

diagnosis and differentiation of pox viruses across different host species, as well as among various strains of the same species (Abdallah and Hassanin, 2013; Gyuranecz *et al.*, 2013). PCR analysis targeting the P4b gene was performed on four FPV and four PPV field samples, and all samples tested positive (100%). The results obtained were similar to those reported by Sultana *et al.* (2019). The present study demonstrated partial sequencing of the 4b core protein gene of four FP and four pigeon virus isolates. The resulting sequences, formatted as FASTA files, were then analyzed using NCBI BLASTN to assess their identity with existing isolates in the NCBI database. To further explore the relationships among these viruses, phylogenetic analyses were performed using maximum likelihood neighbor-joining with MEGA11 software. The analysis revealed that the four FPV isolates in this study exhibited a high degree of similarity to one another, indicating a close genetic relationship, although some samples were frozen many years ago. Similarly, the four-pigeon virus isolates also showed significant homogeneity within their groups, and no molecular characterization study of the FPV in Libya. The FPV isolated in this study were 100% identical to selected isolates from various countries, including Iraq, Iran, India, Brazil, South Africa, Gabon, and Egypt. The Iraqi isolate was described by Hasan *et al.* (2021),

Table 6. Comparison of Libyan pigeon pox isolates with those from other populations (BLASTN).

Identity (%)	Origin of isolate (country/city)	Year of submission	Accession number	Isolate definition
100	Egypt / Giza	2024	PP537572	PPV isolate p/1-2 core protein (P4b) gene.
100	Egypt / Giza	2024	PP537573	PPV isolate p/1-3 core protein (P4b) gene.
100	India / Meerut	2022	ON854919	PPV isolate 01 Vikas aiswal FPV4b core protein gene.
99.80	Taiwan / Taichung	2023	OR515600	PPV isolate Taichung p4b gene.
99.78	India / Meerut	2022	OP066311	PPV isolate SVPUAT_Meerut_02 P4b core protein (P4b) gene.
99.76	Iraq / Baghdad	2020	MT499381	PPV isolate RSJ10 4b core protein (p4b).
99.61	Turkey / Sanliurfa	2022	ON932087	PPV isolate PPV/Harran/13 virion core protein P4b (HM89_gp173) gene.
99.61	Egypt / Ismailia	2023	OR027034	PPV isolate PP/EG/ISMAILIA-4 P4B gene.
99.61	Iraq / Babil	2023	PP537783	PPV isolate Khitam-W10 virion core protein P4b (P4b)gene.
99.61	Iran / Khorasan Razavi	2017	MF102270	PPV isolate FZRP9C 4b core protein (P4b) gene.
99.61	Gabon / Haut-Ogooue	2022	OQ077517	PPV isolate ow_2017_3 4b core protein gene.
99.60	Canada / Guelph	2018	MH175237	PPV isolate ROPI/W370/ON/2012 4b virion core protein (4b) gene.

who showed that nucleotide sequence alignments revealed three Iranian and one Egyptian virus isolates from chicken hosts that were identical to the Iraqi FPV isolate.

The similarities among the PPVs isolated in this study were 100% identical to those isolated from Egypt and India. Furthermore, our analysis revealed identities of 99.80%, 99.78%, and 99.76% with isolates from Taiwan, India, and Iraq, respectively. These high levels of genetic similarity suggest a close evolutionary relationship among these viruses, which may indicate shared transmission pathways or common ancestral origins. Additionally, our isolates demonstrated 99.61% identity with other isolates from Egypt, Iraq, Iran, and Gabon, highlighting the interconnectedness of PPV strains across these regions. Notably, our isolates also showed 99.60% identity with one isolate from Canada, suggesting that the genetic diversity of PPVs may extend beyond traditional geographic boundaries. This level of genetic congruence suggests potential avenues for understanding the epidemiology and transmission dynamics of the FPV across different

geographic regions. The similarities observed among the PPVs warrant further investigation.

These findings underscore the importance of continued surveillance and genetic analysis of avian viruses globally, as they can provide crucial insights into their epidemiology and potential impacts on avian populations.

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Conflict of interest

Abdulatif Ashag hereby declares that he has no financial, personal, or professional conflicts of interest related to the subject matter of this publication. If any potential conflicts arise in the future, he commits to disclosing them promptly to the relevant parties to

ensure transparency and maintain the integrity of the work. Abdulatif Asheg [Date] 29-3-2025.

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Authors' contributions

Dr. Abdulatif Asheg served as the primary supervisor for this work. Dr. Marwa H.A. Elshwihdi was responsible for sample collection and conducted the laboratory work. Dr. Abdulwahab M. Kammon acted as a co-advisor and was responsible for histopathological readings.

Data availability

The data that support the findings of this study are available upon reasonable request.

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